

***Leishmania major*, Strain NIH SD
(MHOM/SN/74/SD)**

Catalog No. NR-42488

For research use only. Not for human use.

Contributor:

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Manufacturer:

BEI Resources

Product Description:

Protozoa Classification: *Trypanosomatidae*, *Leishmania*

Species: *Leishmania major*

Subgenera: *Leishmania*

Strain: NIH SD (MHOM/SN/74/SD) [also known as strain
NIH S (MHOM/SN/74/Seidman) and (MHOM/SN/74/SD)]^{1,2}

Original Source: *Leishmania major* (*L. major*), strain NIH SD
(MHOM/SN/74/SD) was isolated in 1973 from a human
patient with cutaneous leishmaniasis in Senegal, West
Africa.²⁻⁴

Comment: *L. major*, strain NIH SD (MHOM/SN/74/SD) is a
wild-type strain within the *Leishmania* subgenus and is
used in the targeted deletion of leishmanolysin (*gp63*), a 63
kDa proteinase involved in the interaction of promastigotes
and host macrophage receptors and evasion of the
complement system.²

Leishmaniasis is caused by parasitic protozoa of the genus
Leishmania, which is transmitted to both humans and
animals by female phlebotomine sandflies.^{5,6} The sandflies
inject the infective stage (promastigotes) of the parasite from
their proboscis. Promastigotes that reach the puncture
wound are phagocytized by macrophages and other types of
mononuclear phagocytic cells. Inside the cells promastigotes
transform into the tissue stage of the parasite (amastigotes)
and multiply by simple division and infect other mononuclear
phagocytic cells. Infection is endemic throughout the tropics,
subtropics, and Mediterranean basin.^{5,6}

The current taxonomic classification includes two subgenera,
Leishmania, which are found in the midgut of the vector's
intestine, and *Viannia*, which are found in the hindgut of the
vector's intestine. Additionally, the more than 30 known
species of *Leishmania* are divided into New World and Old
World species, whose divergence is thought to correspond to
the separation of the continents millions of years ago. The
subgenera *Leishmania* is comprised of New and Old World
species while the subgenera *Viannia* is comprised of only
New World species.^{7,8} Pathogenic species of both
subgenera have also been grouped into complexes based on
phylogenetic analyses.⁹

Material Provided:

Each vial of NR-42488 contains approximately 0.5 mL of
culture in 10% glycerol. Please see Appendix I for
cryopreservation instructions.

Packaging/Storage:

NR-42488 was packaged aseptically in screw-capped plastic
cryovials and is provided frozen on dry ice. The product
should be stored at cryogenic temperature (-130°C or
colder), preferably in the vapor phase of a liquid nitrogen
freezer. If liquid nitrogen storage facilities are not available,
frozen cryovials may be stored at -70°C or colder for
approximately one week. Note: Do not under any
circumstances store vials at temperatures warmer
than -70°C. Storage under these conditions will result in the
death of the culture.

To insure the highest level of viability, the culture should be
initiated immediately upon receipt. Any warming of the
product during shipping and transfer must be avoided, as this
will adversely affect the viability of the product. For transfer
between freezers and for shipping, the product may be
placed on dry ice for brief periods, although use of a portable
liquid nitrogen carrier is preferred. Please read the following
recommendations prior to using this material.

Growth Conditions:

Modified M199 medium (ATCC® medium 2736), adjusted to
contain 10% (v/v) heat-inactivated fetal bovine serum
(HIFBS) and 10 µg/mL hemin

Incubation:

Temperature: 25°C

Atmosphere: Aerobic

Propagation:

1. Place the frozen vial in a 35°C to 37°C water bath and
thaw for approximately 2 to 3 minutes. Immerse the vial
just enough to cover the frozen material. Do not agitate
the vial. Do not leave the vial in the water bath after it is
thawed.
3. Immediately after thawing, aseptically transfer the
contents of the vial to a T-25 tissue culture flask
containing 10 mL Modified M199 medium.
4. Screw the cap on tightly and incubate the tube or flask at
25°C.

Maintenance:

1. When the culture is at or near peak density, transfer
approximately 0.1 to 0.2 mL into to a new flask
containing 5 to 10 mL fresh Modified M199 medium.
2. Screw the caps on tightly and incubate at 25°C.
3. Transfer the culture every 7 to 14 days as described in
Maintenance steps 1 and 2. The transfer interval will
depend on the size of the inoculum and the quality of the
medium. This should be determined empirically by
examining the culture on a daily basis until conditions for
stable growth have been achieved. Do not allow the
culture to overgrow. Viability of the culture may be
affected soon after reaching peak density.

Please see Appendix I below for cryopreservation instructions.

Citation:

Acknowledgment for publications should read “The following reagent was obtained through BEI Resources, NIAID, NIH: *Leishmania major*, Strain NIH SD (MHOM/SN/74/SD), NR-42488.”

Biosafety Level: 2

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the following publication: U.S. Department of Health and Human Services, Public Health Service, Centers for Disease Control and Prevention, and National Institutes of Health. Biosafety in Microbiological and Biomedical Laboratories. 5th ed. Washington, DC: U.S. Government Printing Office, 2009; see www.cdc.gov/biosafety/publications/bmbl5/index.htm.

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References:

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Dendritic Cells for CD40 Ligand-Induced Interleukin-12p70 Secretion is Strain and Species Dependent.” Infect. Immun. 70 (2002): 3994-4001. PubMed: 12117904.

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4. Neva, F. A., D. Wyler, and T. Nash. “Cutaneous Leishmaniasis – A Case with Persistent Organisms after Treatment in Presence of Normal Immune Response.” Am. J. Trop. Med. Hyg. 28 (1979): 467-471. PubMed: 222157.

5. Chappuis, F., et al. “Visceral Leishmaniasis: What are the Needs for Diagnosis, Treatment and Control?” Nat. Rev. Microbiol. 5 (2007): 873-882. PubMed: 17938629.

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7. Schönian, G., E. Cupolillo and I. Mauricio. “Molecular Evolution and Phylogeny of *Leishmania*.” Drug Resistance in Leishmania Parasites: Consequences, Molecular Mechanisms and Possible Treatments. Eds. A. Ponte-Sucre, E. Diaz, and M. Padrón-Nieves. Vienna: Springer, 2013. 15-44.

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APPENDIX I: CRYOPRESERVATION

1. To harvest the *Leishmania* culture, remove the media containing promastigotes from infected culture flasks that have reached peak density and transfer to 15 mL plastic centrifuge tubes. Centrifuge at 800 x g for 10 min.
2. Remove all but 0.5 mL of the supernatant from each tube, resuspend the cell pellets, and pool them to a single tube.
3. Adjust the cell concentration to 2×10^7 to 4×10^7 cells/mL with fresh Modified M199 medium.
Note: If the concentration of cells is too low, centrifuge at 800 x g for 10 minutes and resuspend in a smaller volume of fresh medium to yield the desired parasite concentration.
4. Mix equal volumes of parasite suspension and fresh medium containing 10% DMSO to yield a final concentration of 1×10^7 to 2×10^7 cells/mL in 5% DMSO. The freezing process should start 15 to 30 minutes following the addition of cryoprotective solution to the cell suspension. Note: To prevent culture contamination, penicillin-streptomycin solution (ATCC[®] 30-2300) may be added to a final concentration of 50 to 100 IU/mL penicillin and 50 to 100 µg/mL streptomycin.
5. Dispense 0.5 mL aliquots into 1 to 2 mL sterile plastic screw-capped vials for cryopreservation.
6. Place the vials in a controlled rate freezing unit. From room temperature cool the vials at $-1^\circ\text{C}/\text{min}$ to -40°C . If the freezing unit can compensate for the heat of fusion, maintain rate at $-1^\circ\text{C}/\text{min}$ through this phase. At -40°C , plunge vials into liquid nitrogen. Alternatively, place the vials in a Nalgene 1°C freezing container. Place the container at -80°C for 1.5 to 2 hours and then plunge vials into liquid nitrogen.
7. Store in either the vapor or liquid phase of a nitrogen refrigerator (-130°C or colder).